

10. MULTI-PHOTON MICROSCOPY

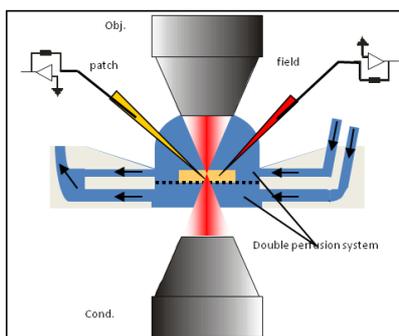
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IN VITRO 2D TWO-PHOTON MEASUREMENT OF NETWORK ACTIVITY (PPKE)

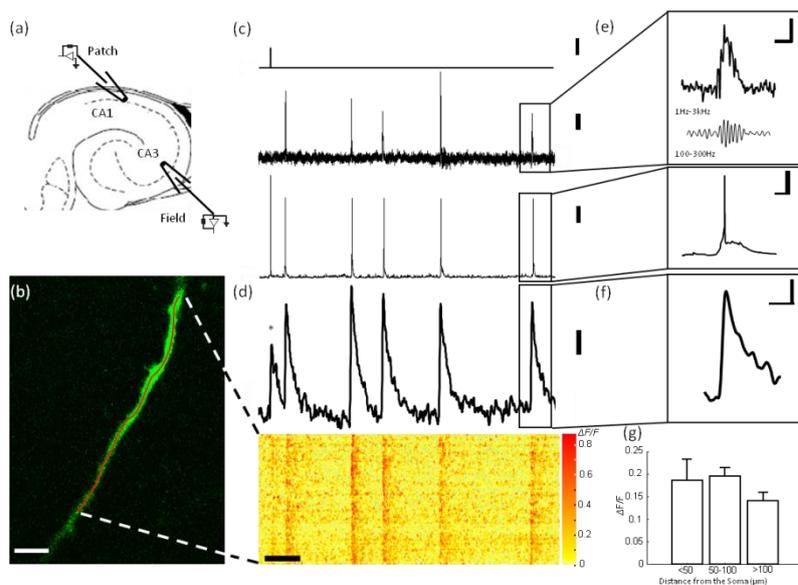
Sharp wave-ripples (SPW-R) activity is involved in the process of memory consolidation. We investigate spontaneous single cell neuronal activities during SPW-R in the hippocampus CA3 region under *in vitro* conditions. Fast spiking (FS), PV+ basket cells (BCs) as the clockworks for neuronal oscillations are important elements of hippocampal neuronal networks. Thus, we focus on PV+ interneurons to reveal the dendritic calcium dynamics during SPW-R. Recently we have extended our interest of these measurements with the principal, pyramidal neurons to the better understand of the spontaneous neuronal network activity.

To achieve this, we combine two-photon microscopy, local field electrophysiology, single cell electrophysiology, and dendritic patch clamp recordings. To measure the pharmacological background of calcium dynamics in single cell dendrites, we use focal synaptic stimulation and two-photon uncaging of novel, own developed glutamate and GABA caged compounds.

1)



2)



3)

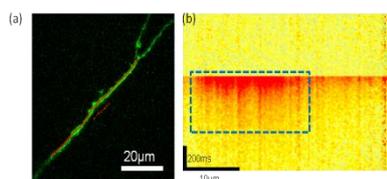


Fig. 1 1) Experimental arrangement to generate spontaneous network activity in *in vitro* conditions. 2) Simultaneous two-photon calcium imaging, whole cell patch clamp recording and local field potential recording in *in vitro* measurement. 3) Two-photon uncaging and calcium imaging. (Chiovini B et al. 2010 *Neurochem Res.*)

IN VIVO 3D TWO-PHOTON MEASUREMENT OF NETWORK ACTIVITY

To understand the fast computational mechanisms of the brain, one needs to be able to perform rapid measurements at several sites along a single neuron as well as to image large populations of neurons. Traditional two-dimensional measurements are severely limited for such kinds of endeavors since neurons are located in three dimensions. To overcome this problem, we have developed new solutions to perform three-dimensional functional imaging with large scanning ranges along the z direction. With our three-dimensional microscopes we are able to maintain random access point scanning with a short pixel dwell time. The speed and scanning volume of our technique in combination with the $\sim 800 \mu\text{m}$ penetration dept of two-photon technology makes our methodology very convenient for *in vivo* measurements of neuronal populations, too.

Populational activity has long been studied in the visual cortex. We conduct *in vivo* two-photon imaging of the V1 area after using multicell bolus loading of a calcium indicator dye. Neuronal network responses are followed after visual stimulation using a moving bar or moving grating protocol. In addition, active cells are selected based on the previously recorded somatic activity and their dendritic responses are followed along with the network activity in three dimensions by using whole-cell patch clamp techniques.

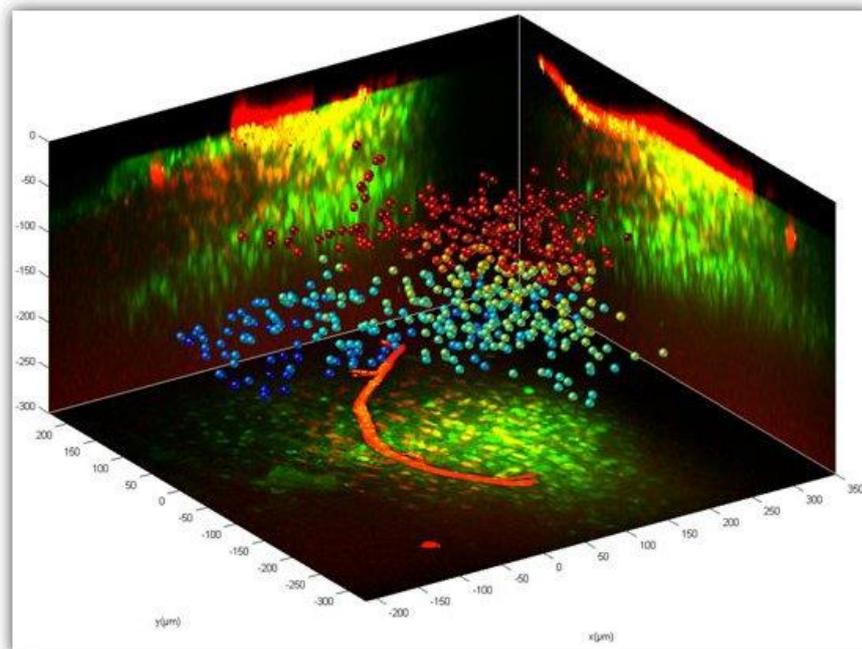


Fig. 2 Projection of the 3D volume with multicell bolus loading of a calcium indicator dye. Points represents 251 simultaneously measured cells.

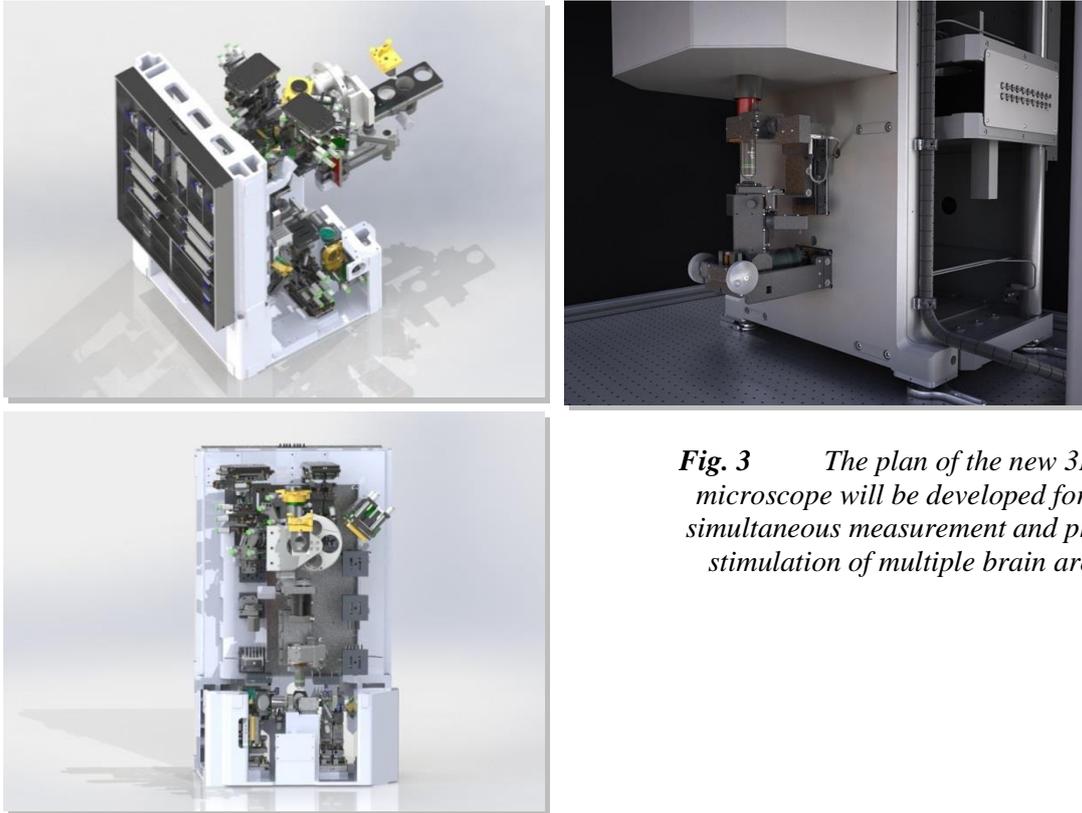


Fig. 3 The plan of the new 3D microscope will be developed for the simultaneous measurement and photo-stimulation of multiple brain areas

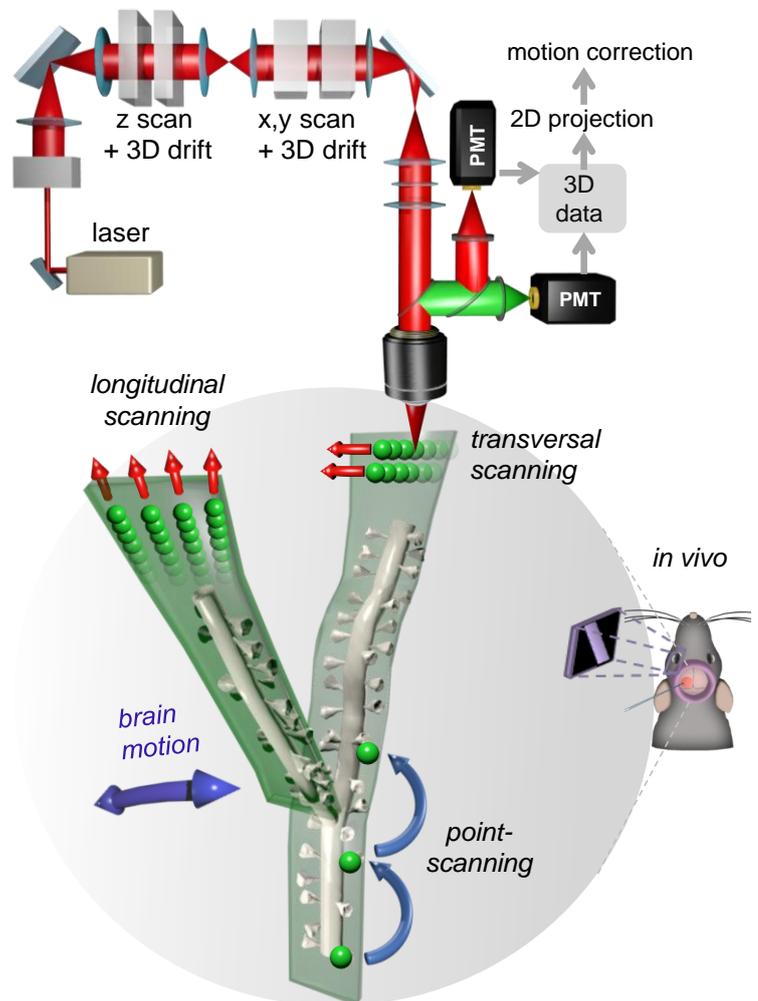


Fig. 4 New microscope technologies were developed for the 3D measurements of the moving brain of behaving animals.

PUBLICATIONS

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